

A lineage-specific mitosomal membrane protein possibly involved in vacuole-mitosome contact in *Entamoeba histolytica*

Herbert J. Santos^{1,2}, Yuki Hanadate^{1,3}, Kenichiro Imai⁴, and Tomoyoshi Nozaki^{1,2}

¹ Department of Parasitology, National Institute of Infectious Diseases, Japan.

² Department of Biomedical Chemistry, Graduate School of Medicine, The University of Tokyo, Japan. ³ Graduate School of Life and Environmental Sciences, University of Tsukuba, Ibaraki, Japan. ⁴ Biotechnology Research Institute for Drug Discovery, National Institute of Advanced Industrial Science and Technology, Tokyo, Japan.

The anaerobic protozoan parasite *Entamoeba histolytica* is the etiological agent of amoebiasis, a disease characterized by diarrhea and dysentery, which could lead to extra-intestinal abscess formation in the liver, lungs, and brain. This organism lacks mitochondria and instead possesses a highly degenerate and divergent mitochondrion-related organelle (MRO) called mitosome. Mitochondria and MROs are bound by a double-membrane barrier, lined with various proteins that allow for protein and metabolite transport, lipid transfer, signaling, dynamics and inter- and intra-organelle crosstalk. However, only a few protein homologs have been identified in the outer and inner membranes of the *E. histolytica* mitosome, suggesting that the mitosomal membranes are equipped with components that may have been modified by unique and possibly lineage-specific elements. Using a specialized prediction pipeline, we searched for proteins possessing α -helical transmembrane domains that are unique to *E. histolytica* mitosomes. A total of 25 candidate proteins emerged and following screening by localization analysis, we confirmed three novel *Entamoeba*-specific transmembrane mitosomal proteins (ETMPs). ETMP1 was predicted to have one transmembrane domain and two coiled-coil regions by *in silico* analyses. It was demonstrated to be integrated to the mitosomal membranes based on carbonate fractionation and immunogold analysis. ETMP1 forms a 180 kDa complex and immunoprecipitation analysis detected a candidate interacting partner that is localized to vacuolar and vesicular membranes. We expressed this ETMP1-binding partner and subsequent immunofluorescence and immunoelectron microscopy data demonstrated an unprecedented vacuole-mitosome contact.